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Investigation of Coenzyme Q₁₀ by Voltammetry

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Abstract

A simple, accurate and rapid voltammetric method has been developed for the quantitative determination of coenzyme Q_{10} . Studies with direct current voltammetry were carried out using a glassy carbon electrode (GCE) in a phosphate buffer solution (pH 6.86). A well-defined oxidation peak of Co Q_{10} was obtained at -0.600 V vs Ag/AgCl. The magnitude of the oxidation peak current has been found to be related to the concentration of the coenzyme over the range of (2.0·10⁻⁵ to 2.0·10⁻⁴ M) (r = 0,991). Antioxidant activity of Co Q_{10} was investigated.

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1. Introduction

Coenzyme Q_{10} (2,3-dimetoxy-5-methyl-6-decaprenyl-1,4-benzoquinone, CoQ_{10}) is a fat soluble, vitamin-like quinone commonly known as ubiquinone or ubidecarenone. CoQ_{10} (fig. 1) is widely distributed in the living system of plants, animals and humans¹. It is essential for cell respiration and electron transfer and it acts as an electron carrier and translocator of hydrogen ions in mitochondria². Coenzyme Q_{10} exhibits antioxidative and prooxidative properties³.

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Fig. 1. Structural formula of coenzyme Q₁₀

The earliest observations of an antioxidant function of $CoQ_{10}H_2$ (fig. 2) date back to the sixties, and show that this substance could, like α -tocopherol, protect against the light-catalysed prooxidation of mitochondrial lipids. These observations have been extensively confirmed during the last two decades. Coenzyme QH_2 is the only lipid-soluble antioxidant synthesised by animal tissues *in vivo*. Coenzyme $Q_{10}H_2$ can act as an antioxidant directly, by preventing initiation and propagation of mitochondrial lipid peroxidation, or indirectly, by recycling α -tocopherol⁴. Coenzyme $Q_{10}H_2$ can also protect against oxidative damage to other important cellular macromolecules such as proteins and DNA.

Fig. 2. Structural formula of ubiquinol CoQ10H2

The reactivity of $CoQ_{10}H_2$ with peroxyl radicals is much slower than that of α -tocopherol, and because $CoQ_{10}H_2$ is a highly hydrophobic molecule located in the middle of the phospholipid bilayer, it has considerably less intramembrane mobility than α -tocopherol, which would tend to decrease its radical scavenging potential. Therefore, this suggests that α -tocopherol, rather than $CoQ_{10}H_2$, acts as a direct scavenger of peroxyl radicals within the mitochondrial inner membrane.

Coenzyme Q_{10} is an effective scavenger of free radicals and takes part in the elimination process of products resulting from metabolic reactions. Both CoQ_{10} (CoQ_{10} and $CoQ_{10}H_2$) and vitamin E efficiently detoxify cells from toxins. The coenzyme also prevents cells and membranes not only against generating free radicals but also from oxidative modifications of proteins, lipids and DNA⁵.

The anodic oxidation of coenzyme Q_{10} on a glassy carbon electrode (GSE) in a phosphate buffer solution (pH=6.86) was investigated. Considering obtained results, a simple, reproducible, rapid and accurate method was developed to determine CoQ_{10} .

2. Materials and Methods

Chemicals used were coenzyme Q_{10} (Co Q_{10}), concentration > 98%, (Sigma-Aldrich).

Standard solution for electrochemical experiments was prepared by dissolving a suitable amount of coenzyme Q_{10} in an ethyl alcohol followed by heating to a temperature not exceeding 25^oC. The stock solution was stored in the dark and cool. Background solution was prepared by dissolving standard powder of Na₂HPO₄ in 1 L volumetric flask.

All voltammetric experiments were performed using an electrochemical analyzer "TA-2" ("Tomanalyt", Tomsk) with a three-electrode cell connected to it in which the glassy carbon electrode of 1 mm in diameter and silver chloride electrode were used as a process electrode and reference electrode, respectively. All potentials were measured and reported on the external silver chloride electrode with 1M NaCl solution. A method of determining electrochemical properties was made by cyclic voltammograms without addition and with following addition of the

test substances in a supporting electrolyte solution. The range of potential was from -1.5 to 1.5 V. Oxygen was removed by bubbling nitrogen through the electrochemical cell.

Antioxidant Activity Estimation

The antioxidant activity was estimated using an electrochemical method. The experimental method consisted in the recording of a voltammogram of the cathode electroreduction of oxygen by means of AOA Analyzer (RU.C.31.113.A N28715) connected to a PC. The antioxidant activity of the tested samples was calculated according to kinetic criterion K (in micromoles per liter minute) indicating the quantity of the reactive oxygen species in time and determined using the formula [6]:

$$K = \frac{C_o}{t} \left(1 - \frac{I}{I_o}\right)$$

where

C₀₂ - oxygen concentration in the initial solution without a substance, in micromoles per liter;

 I_i - electric current of oxygen electroreduction, in microampere;

 I_o - electric current of oxygen electroreduction without a substance in the solution, in microampere;

t - process running time, in minute.

3. Results and Discussion

Initially the effect of the supporting electrolyte pH on the analytical signal of coenzyme Q10 at different concentrations in solution was considered. As a result the cyclic voltammogram with redox peaks of coenzyme Q10 on the GCE at a scan rate 30 mV/s was obtained (Figure 3).



Fig. 3. Cyclic voltammogram of CoQ_{10} on GCE ($\nu = 30 \text{ mV/s}$) at different concentration CoQ_{10} in the solution: (1) background electrolyte without CoQ_{10} ; (2) $0.5 \cdot 10^4 \text{ M } CoQ_{10}$; (3) $1.0 \cdot 10^4 \text{ M } CoQ_{10}$; (4) $1.5 \cdot 10^4 \text{ M } CoQ_{10}$; (5) $2.0 \cdot 10^4 \text{ M } CoQ_{10}$ in the phosphate buffer solution (pH=6.86).



The influence of pH supporting electrolyte on redox signal coenzyme Q_{10} on GCE was investigated (fig. 4).

Fig. 4. Dependence of the oxidation current (A) and reduction current (B) of coenzyme Q10 on the GCE from their concentration in the solution at different pH value: 1.65 (1); 4.01 (2); 6.86 (3); 9.18 (4).

With addition of the analyte in the acidic and alkaline buffer lower values of the analytical signal in the anode and cathode area, respectively, are observed. It is assumed that protonation of coenzyme Q_{10} molecule occurring in acidic medium facilitates its oxidation. In alkaline medium analytical signal due to complete dissociation of hydroxyl groups (-OH) of hydroquinone group of coenzyme Q_{10} molecule increases and as a result facilitates the recovery process.

For the future investigation as a supporting electrolyte the phosphate buffer solution with pH 6.86 was chosen. This caused the stability of the hydroquinone group of coenzyme Q_{10} in a neutral supporting solution.

On the basis of obtained data and literature [7] a redox mechanism of coenzyme Q_{10} on GCE can be suggested. In aqueous medium at neutral pH, the reduction is either by two protons two electrons (fig. 5).



Fig. 5. Redox scheme of CoQ₁₀ on GCE in aqueous medium at neutral pH.

On a glassy carbon electrode, at pH 6.86 coenzyme Q_{10} shows two sets of waves: one at a positive (+ 0.4 V) potential and another at a negative (- 0.6 V) one. The oxidation peak at + 0.4 V is reversible and involves the oxidation of the hydroquinone group. The reduction peak at negative potential of -0.6 V was similar to the previously reported reduction of ubiquinon to ubiquinol.

The calibration curve (fig. 6) of the peak current versus concentration is constructed using the linear regression method. The calibration curves are linear within the range of concentration from 2.0×10^{-5} to 2.0×10^{-4} M. This range of concentration is available for determination coenzyme Q₁₀ in pharmaceutical and cosmetics objects.



Fig. 6. Calibration curve of the redox peak current versus concentration coenzyme Q_{10} (oxidation (A) and reduction (B))

Also the antioxidant activity of coenzyme Q_{10} has been investigated. Results are shown in Table 1.

$C \times 10^5$, M	K, mkmol/(L•min)
0.1	0.32±0.09
1.0	0.25±0.05
10.0	0.18±0.06

Table 1. Values of the antioxidant activity at various concentrations of coenzyme Q10 in the solution

As it is seen from the table 1 the activity of coenzyme Q_{10} decreases with increasing concentration of the object in a solution. However, the change of coenzyme Q_{10} antioxidant activity does not strongly depend on its concentration.

4. Conclusion

As results electrochemical properties of coenzyme Q_{10} by cyclic voltammetry were investigated. The redox mechanism of coenzyme Q10 on GCE in aqueous solution has been assumed. The range of linear dependence of the analytical signal height on its concentration in the solution has been found. This method is suitable for determination of CoQ_{10} in pharmaceutics. The main advantages of this method are significantly shortening of analysis time, low cost of analysis and widespread access to apparatus.

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